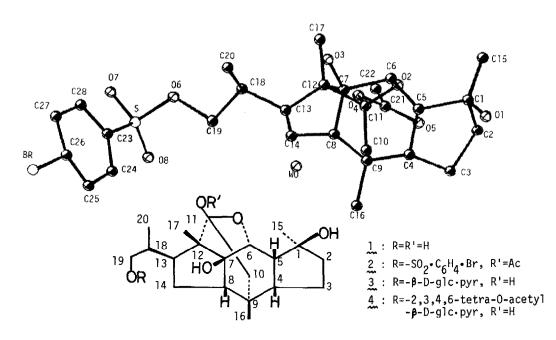
CINNCASSIOL D1 AND ITS GLUCOSIDE, NOVEL PENTACYCLIC DITERPENES FROM CINNAMOMI CORTEX

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Summary : A new pentacyclic diterpene with a new skeleton and its glucoside were obtained from the fraction exhibiting anti-complement activity of the water extractive of Cinnamomi Cortex. Their structures were elucidated as 1 and 3 by chemical, spectral and X-ray crystallographic studies.

Two new pentacyclic diterpenes, named cinncassiol D_1 (1) and cinncassiol D_1 glucoside (3), were isolated from the fraction exhibiting anti-complement activity¹⁾ of the water extractive of Cinnamomi Cortex ("Tōkō Keihi"; the dried bark of <u>Cinnamomum cassia</u> Blume).

Cinncassiol D₁ (1), a white powder, $[r]_{D}$ +12.7° (MeOH), $C_{20}H_{32}O_{5}$ (FD-MS (m/z) : 352 (M⁺), 335 (M⁺-OH); EI-MS (m/z) : 352.220 (M⁺)), showed in the IR spectrum the absorption due to hydroxyl (3440 cm⁻¹). NMR spectra (in Py-d₅) are as follows; ¹³C **s**: 10.6, 13.5, 24.4, 28.1 (4 x CH₃-), 25.2, 25.5, 37.3, 40.4 (4 x -CH₂-), 36.9, 40.4, 48.6, 51.0, 54.1 (5 x >CH-), 41.0, 57.1 (2 x >C<), 67.3 $(-CH_2-0-)$, 77.2 (>CH-0-), 82.0, 88.6 (2 x \geq C-0-) and 107.5 ($>C \leq_{0-}^{0-}$), ¹H **s**: 0.90, 1.69, 1.69 (3H each, all s, 3 x tert.CH₃), 1.37 (3H, d, J=7 Hz, sec.CH₃), 3.80 (2H, d, J=8 Hz, -CH₂-O-), 4.44 (1H, br. s, >CH-CH-O-). Based on the above spectral data, <u>1</u> was supposed to be a new diterpene. <u>]</u> was treated with p-bromobenzenesulfonyl chloride and Py and the product was acetylated (Ac20-Py, overnight at r.t.) to give the monoacetyl monobrosylate (2), colorless plates, mp 104-105°, $[\mathbf{a}]_{D}$ 0° (MeOH), ¹H-NMR (in CDC1₃) **5** : 2.03 (3H, s, -OAc), 7.69, 7.81 (2H each, double d of J=10 Hz, 4 x arom. A single crystal of 2 suitable for a X-ray diffraction study was obtained by recrystalliproton). zation from dil.MeOH and its data are as follows; $C_{28}H_{37}SO_8Br \cdot H_2O$; monoclinic, space group P2₁ (Z= 2); lattice constants <u>a</u>=10.222(6), <u>b</u>=10.271(7), <u>c</u>=15.155(6) Å, **β**=96.91(4)°, V=1579.6 Å³; D(calcd.)= 1.33, D(obsd.)=1.37 g/cm³. The intensity data of 2192 (26 \leq 45°) were measured with a Syntex R₃ full automated four-circle diffractometer using $w-2\theta$ scan technique and graphite-monochromated Mo (Ka) radiation (A=0.71069 Å). The structure was solved by heavy atom method. Block diagonal least squares refinements with isotropic nonhydrogen atoms have currently converged to a standard residual of 0.105 for the 1284 obseved reflections (I \geq 2.0 σ (I)). A computer-generated perspective drawing thus obtained is shown in Fig. 1. Therefore, the molecular structure of cinncassiol D₁ is represented by the formula <u>l</u>or its enantiomer.



Cinncassiol D₁ glucoside (3), a white powder, $[\mathbf{A}]_D$ -4.1° (MeOH), $C_{26}H_{42}O_{10}$ (FD-MS (m/z) : 553 (M + K⁺), 537 (M + Na⁺)), on enzymatic hydrolysis with crude hesperidinase liberated cinncassiol D₁ (1) and D-glucose. While 3 was acetylated (Ac₂O-Py, for 20 min. at r.t.)²⁾ to yield the tetraacetate (4), a white powder, $[\alpha]_D$ -18.6° (MeOH), $c_{34}H_{50}O_{14}$, ¹H-NMR (in CDCl₃) 5: 2.00, 2.03, 2.05 and 2.09 (4 x OAc), 4.49 (1H, d, J=7 Hz, amomeric proton of glucoside), EI-MS (m/z) : 331.102 $(C_{14}H_{19}O_{9}^{+}; terminal peracetylated hexosyl cation).$ The above spectral data for 4 indicate that D-glucosyl moiety is linked to the C-19 hydroxy group³⁾ of 1 and is β -configuration. The structure of 3 was threfore elucidated as cinncassiol D_1 19-0- β -D-glucopyranoside.

They are woth of note as novel type diterpenes (1 and 3) with a new skeleton which are assumed to be the key substances possessing anti-complement activity.

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References and Notes

1) A.Koda, E.Katsuta, S.Watanabe and M.Mizuno, Nippon Yakurigaku Zasshi, 66, 366 (1970).

- 2) On acetylation in the same way, 1 gave 19-0-monoacetyl cinncassiol D_1 . 3) It was also supported from the evidence of ¹³C-NMR spectrum of 4. Co 'Comparison of the ¹³C-NMR spectra of 4 with that of 1 indicated that C-19 signal is shifted downfield by 7.8 ppm and C-18 signal is shifted upfield by 3.1 ppm due to glycosylation shifts (R.Kasai, M.Suzuo, J.Asakawa and O.Tanaka, Tetrahedron Lett., 1977, 175; K.Tori, S.Seo, Y.Yoshimura, H.Arita and Y.Tomita, Tetrahedron Lett., 1977, 179).